

Chemical Engineering Thesis Defense

Engineering and Investigating the Effects of
Renewable Chemical Production in Bacteria

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Abstract

Metabolic engineering of bacteria has become a viable technique as a sustainable and efficient method for the production of biochemicals. In the first study, genes that were shown to be highly differentially expressed when exposed to styrene were investigated in *E. coli* by testing their deletion and overexpression strains. It was found that *plsX*, a gene responsible for the phospholipid formation in membranes, had the most promising results when overexpressed at 10 μM IPTG, with a relative OD600 of $706 \pm 117\%$ at 175 mg/L styrene when compared to the control plasmid at the same concentration. This gene is likely to be an effective target when engineering styrene- and other aromatic-producing strains, increasing titers by reducing the cytotoxicity. In the second study, cyanobacterium *Synechococcus* sp. PCC 7002 was being engineered for the overproduction of L-serine. As a robust, photosynthetic bacteria, it has potential for being used in such-rich regions to capture CO₂ and produce industrially relevant products. In order to increase L-serine titers, a key degradation gene, *ilvA*, was explored. While *ilvA* is responsible for degrading L-serine into pyruvate, it is also responsible for initiating the only known pathway for the production of isoleucine in PCC 7002. Herein, a plasmid containing the native A0730 gene was designed to be transformed into a strain of *E. coli* with an *ilvA* knockout to investigate its potential to restore isoleucine production. If shown to be effective, a *Synechococcus* sp. PCC 7002 $\Delta ilvA$ strain can be engineered with minimal effects on growth and is expected to lead to increased L-serine accumulation.



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Zoom Link: <https://asu.zoom.us/j/85668516273>